

Figure 1. Stern-Volmer plot for the quenching of 1-cyanonaphthalene fluorescence vs. [H⁺]. The inset shows the region of 0-1.5 M expanded.

 Table I. Singlet Lifetimes and Stern-Volmer Results for the Naphthalene Derivatives 2-7

compd	$ au,^a$ ns	$k_{q} au, {}^{b,c}M^{-1}$	10 ⁻⁹ k _q , M ⁻¹ s ⁻¹	$k_{q} au, \overset{c,d}{M^{-1}}$	$10^{-7}k_q, M^{-1}s^{-1}$
2 ^e	9.28	11.4 ± 0.2	1.22		
3	9.88	26.0 ± 0.1	2.63		
4	8.96	12.0 ± 0.07	1.36		
5 ⁄	10.5	<0.1 ^g	<0.01	0.072 ± 0.001	0.68
6	14.5	0.28 ± 0.02^{g}	0.019	0.9991(6) 0.292 ± 0.002	2.0
7	9.86	0.98(7) 0.22 ± 0.01^{g}	0.017	0.9998(5) 0.216 ± 0.005	2.2
		0.98 (5)	<u>-</u>	0.998 (6)	

^aBy single photon counting. ^bUsing [H⁺] by titration. ^cErrors are standard deviation of the least-squares slope with correlation coefficient and number of points given below. ^dUsing [H⁺]^{hv}. ^eValues of 8.9 ns, 9.6 M⁻¹, and 1.08 × 10⁹ M⁻¹ s⁻¹ have been reported for 20% acetonitrile/80% water.⁹ ^fValues of 10 ns, <0.03 M⁻¹, and <3 × 10⁶ M⁻¹ s⁻¹ have been reported.⁹ ^gCalculated from points up to [H⁺] = 2.0 M only.

Finally, some comment about the proton quenching of these substrates is required. The singlet lifetimes for the compounds allow calculations of the k_q values shown in the table. Compounds 4 and 7 are the naphthalene analogues of the benzyl alcohols studied by Wan and Turro.² There is clearly no meta effect. However, the reason is obvious. The 1-methoxy compounds all have similar reactivity with protons and it is well-established that the pathway for quenching of the excited state of 1-methoxy-naphthalene (2) is via ring protonation.⁹ Likely, the others are similarly quenched by protons. Since the 2-methoxy compounds are also all quenched at similar rates (about a factor of 10^2 slower than the 1-methoxy isomers) the most likely process is, again, ring protonation.¹⁷ MO calculations⁹ of the excited state of 1- and

2-methoxynaphthalene support the idea that electron density increases in the ring are greater for the 1-isomer. Clearly, substrates that have several basic centers must be examined quite carefully to determine what the k_q value obtained from fluorescence quenching means in terms of the reactivity of any given functional group. To have ascribed the k_q solely to benzylic cleavage in this case would have been wrong.

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Acrolein Acetals as Allyl Cation Precursors in the Ionic Diels-Alder Reaction

Paul G. Gassman,* Daniel A. Singleton,¹ James J. Wilwerding,² and Subhash P. Chavan

> Department of Chemistry, University of Minnesota Minneapolis, Minnesota 55455 Received November 19, 1986

Diels-Alder reactions that employ acrolein as the dienophile often give undesirable results due to the ease of polymerization of acrolein. In order to circumvent this problem, a variety of Lewis acids³ and high pressures⁴ have been employed. Even under these conditions yields are often less than 50%. Our experience with the ionic Diels-Alder reaction of allyl cations⁵ suggested that acetals of acrolein⁶ (1) should be convenient precursors of the alkoxy-substituted allyl cations **2**, which, in turn, should be ex-

⁽¹⁷⁾ Note that 1-cyanonaphthalene and 2-cyanonaphthalene have very similar k_q values,⁹ suggesting that quenching by protons occurs at nitrogen. The methoxynaphthalenes, in contrast, protonate at ring carbons. This suggests that $[H^{+}]^m$ may be a fundamental property of the medium and not just a method of linearizing Stern-Volmer plots.

⁽¹⁾ National Science Foundation Fellow, 1983-1986.

⁽²⁾ Petroleum Research Fund Undergraduate Summer Fellow, 1986.

⁽³⁾ For typical examples, see: Trost, B. M.; O'Krongly, D.; Belletire, J. L. J. Am. Chem. Soc. 1980, 102, 7595. Danishefsky, S.; Bednarski, M. Tetrahedron Lett. 1985, 26, 2507. Laszlo, P.; Lucchetti, J. Ibid. 1984, 25, 2147.

⁽⁴⁾ Dauben, W. G.; Krabbenhoft, H. O. J. Org. Chem. 1977, 42, 282.
(5) Gassman, P. G.; Singleton, D. A. J. Am. Chem. Soc. 1984, 106, 6085,
7993. Gassman, P. G.; Singleton, D. A. J. Org. Chem. 1986, 51, 3075.

^{(6) 3,3-}Diethoxypropene (acrolein diethyl acetal) was purchased from Aldrich Chemical Co. and used without purification. 2-Vinyl-1,3-dioxolane was prepared according to the literature procedure (Piasecki, A.; Burczyk, B. J. Prakt. Chem. 1985, 327, 543).



Table I. Yields Obtained in the Reactions of Acrolein Acetals with 1.3-Dienes

		R = CH(OC ₂ H ₅) ₂ 1a	R = cH	
diene	product	isolated yield, %	isolated yield, %	GLC yield, %
	₽ R H	57	94	100
\bigcirc	R H	31	63ª	72ª
Ľ	\mathcal{Q}_{R}	48	58 ^b	72 ^b
X		52	65	69
↓°	Ũ,	27	62	63
Ţ		61	56	77

^aThis material is a 12:1 mixture of stereoisomers as determined by NMR analysis. ^bThis material is an 8:1 mixture of stereoisomers as determined by NMR analysis. None of the regioisomer was detected. ^cA 4:1 ratio of isoprene to 1b was used due to the ease of polymerization of isoprene.

cellent dienophiles (Scheme I). We now wish to report studies that confirm this hypothesis.

In a typical procedure utilizing 3,3-diethoxypropene⁶ (1a, R = C_2H_5), 10 mmol of the appropriate diene and 5 mmol of 1a in 50 mL of methylene chloride were cooled to -78 °C and 0.1 mmol (2 mol %) of triflic acid in 1,1,2-trichloro-1,2,2-trifluoroethane was added. The reaction was allowed to warm slowly while the presence of 1a was monitored by GLC. When 1a was no longer detected, 0.1 mL of triethylamine was added, the solvent was removed, and the product was isolated by column chromatography or by medium-pressure liquid chromatography. Table I lists the yields of Diels-Alder adducts obtained with a series of six common dienes.⁷

Scheme II



The cycloaddition of 1a to a series of dienes via the intermediacy of 2a is an attractive reaction because it occurs readily below 0 °C. This can be contrasted with the related Diels-Alder reactions of the intact acetals of α,β -unsaturated ketones and aldehydes, which require long reaction times at high temperature.⁸ As can be noted from Table I, the yields obtained in the addition of 3,3-diethoxypropene were not attractive from a synthetic point of view. We reasoned that a possible cause of these mediocre yields was the diffusion of the alcohol away from the cation 3a with resultant failure to recombine to yield 4a. In order to circumvent this problem, we chose to use 2-vinyl-1,3-dioxolane⁶ (1b, R = $-CH_2CH_2$) as the precursor of 2b and 3b.⁹ In this case, 3b would



<u>3b</u>

have the alcohol moiety tethered and held in the proximity of the resulting carbocation. The data shown in Table I illustrate that in most cases this provided a dramatic increase in the yields of the cycloadducts. Because the isolated yields of products from 2-vinyl-1,3-dioxolane were based on reactions run on a 0.5-g scale, GLC yields were also obtained. These ranged from 63% to 100%.

In order to demonstrate the general scope of this cycloaddition reaction, we examined the effect of methyl substitution on 2vinyl-1,3-dioxolane. Treatment of 1,3-cyclohexadiene with 2methyl-2-vinyl-1,3-dioxolane (5; ethylene glycol acetal of methyl vinyl ketone¹⁰), 2-(2-propenyl)-1,3-dioxolane (6),¹¹ and 2-(1propenyl)-1,3-dioxolane (7),¹¹ gave 8¹² (78% yield; isolated from an 11:1 mixture¹³ of 8 and its anti isomer), 9 (57% yield of a 6.5:1.0

(10) Hahn, E. J. Org. Chem. 1973, 38, 2092.
(11) Acetals 6 and 7 were prepared according to a modification of the published procedure (Piasecki, A.; Burczyk, B. J. Prakt. Chem. 1985, 327, 543).

⁽⁷⁾ Satisfactory elemental analyses and/or exact mass molecular weights were obtained on all new compounds. All new compounds had ¹H NMR, ¹³C NMR, and IR spectra consistent with the assigned structures.

⁽⁸⁾ For examples of intramolecular Diels-Alder reactions of acetals of α , β -unsaturated ketones at temperatures of 170-200 °C for 24-15 h, see: Jung, M. E.; Halweg, K. M. Tetrahedron Lett. 1981, 22, 3929. Bal, S. A.; Helquist, P. Ibid. 1981, 22, 3933.

⁽⁹⁾ For 1b, the volume of solvent was reduced to 25 mL.

⁽¹²⁾ The stereochemistry of 8 was established by both chemical and spectroscopic means. The Diels-Alder adduct of 1,3-cyclohexadiene and (Kozlov, N. S.; Raikova, T. S.; Aramo, A. P. Vestsi Akad. Navuk BSSR, Ser. *Khim. Navuk* (2) 1974, 78). Acetal formation using this adduct and ethylene glycol gave 8. Hydrolysis of 8 with aqueous 10% hydrochloric acid gave material identical with the adduct formed by the literature procedure. Eu-(fod)₃ shift-reagent studies on 8 and on its epimer showed the acetal moiety to be syn to the double bond in 8 and anti to the double bond in the epimer of 8.

mixture¹⁴ of 9^{15} and its anti isomer), and 10 (67% yield of a 27:1 mixture of 10¹⁵ and its anti¹⁶ isomer), respectively (Scheme II). The formation of 8-10 demonstrates that a variety of substitution patterns can be tolerated by 1b without major inhibition of the formal 2 + 4 cycloaddition¹⁷ of the corresponding allyl cation to 1,3-cyclohexadiene. This bodes well for the broad generality of this reaction.

In summary, we have developed a very low temperature process for the addition of acetals of acrolein and its derivatives to 1,3dienes. We believe that the underlying mechanistic concept associated with this facile cycloaddition reaction will find widespread usage. We are currently exploring a variety of applications of these principles to other systems.

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(14) The isomer ratio was determined by the use of ¹H NMR studies. (15) The stereochemistries of 9 and 10 were assigned on the basis of difference NOE studies and shift-reagent studies, respectively.

(16) Trace amounts (3-4%) of an impurity could be detected in 10 by ¹H NMR. The peak positions that could be determined for this impurity were consistent with it having the structure of the anti isomer.

(17) It is interesting to note that the structure of 10 establishes the formal suprafacial nature of our cycloaddition reaction.

Biosynthesis of Brevetoxins. Evidence for the Mixed Origin of the Backbone Carbon Chain and the Possible Involvement of Dicarboxylic Acids

Hong-Nong Chou and Yuzuru Shimizu*

Department of Pharmacognosy and Environmental Health Sciences, College of Pharmacy The University of Rhode Island Kingston, Rhode Island 02881 Received December 5, 1986

The toxins found in the extremely deleterious red tide organism Gymnodinium breve (=Ptychodiscus brevis) possess unusual polycyclic ether structures examplified by brevetoxin A $(1)^1$ and brevetoxin B (2).² Although the ring systems of the two toxins differ significantly, the resemblance of the terminal four rings suggests their closely related biosynthesis. The methyl-substituted carbon chain backbone appeared to be biosynthesized by the straightforward elongation of acetate units and the introduction of methyl groups from methionine and/or propionate as seen with the biosynthesis of most polyketides and fatty acid derivatives. However, the feeding experiments conducted by the authors' group³ and Nakanishi's group⁴ using carbon-13-labeled acetates and methionine resulted in inexplicable labeling patterns in brevetoxin B (2), as summarized in Figure 1A. The puzzling finding prompted us to reexamine the entire ¹³C NMR signal assignment⁵ and repeat the experiments. Now we are able to present experimental data to explain what initially appeared to be irrational results.



Figure 1. Structures of brevetoxin A (1) and brevetoxin B (2), the labeling patterns resulted from feeding experiments with carbon-13-labeled precursors (A) and assumed building blocks of 1 (B). The solid lines in (A) indicate the presence of couplings.

First we observed that the prolonged incubation with carbon-13-labeled acetate led to the random but differential labeling of carbons in 2. In attempts to pulse feed acetate by a brief exposure of the organism to a large dose of methyl-13C-labeled acetate, we isolated 2, whose ¹³C NMR spectrum shows carbon-carbon spin-spin couplings with only selected carbons.

In a typical experiment, the organism was grown in NH-15 medium⁶ in Fernbach flasks (2 L \times 10) under 4000-5000-lux illumination. Eleven days after inoculation, each flask was fed with [2-¹³C₁]CH₃COONa (100 mg), chloramphenicol (5 mg), and streptomycin (10 mg). After 48 h, the culture was harvested, and a total of 1.1 mg of pure 2 was isolated according to the previously established procedure.⁷ The ¹³C NMR spectrum of the isolated toxin⁸ showed enrichment with all the carbons which were found to be of acetate methyl origin in the previous experiments, but most significantly, the signals for the carbon fragments C6-C7-C8, C10-C11, C45-C13-C14-C15, C21-C22, C24-C25, C27-C28, C32-C33, and C35-C36 showed distinct splittings due to spin-spin couplings between the adjacent carbons. No such randomization was observed when carboxyl-labeled acetate was fed under a similar condition.⁸ The observed couplings indicate that those carbon fragments were constructed by rather confined carbon-carbon linkage formation between methyl originated carbons during the concentrated pulse feeding. Furthermore, feeding experiments with $[1,2^{-13}C_2]$ acetate indicate that

⁽¹³⁾ The two components of the mixture were separated by chromatography and the isomer ratio was based on the isolated yields of 78% for 8 and 7% for the epimer of 8.

Shimizu, Y.; Chou, H. N.; Bando, H.; Van Duyne, G. D.; Clardy, J. C. J. Am. Chem. Soc. 1986, 108, 514-515.
 Lin, Y. Y.; Risk, M.; Ray, S. M.; Van Engen, D.; Clardy, J.; Golik, J.; James, J. C.; Nakanishi, K. J. Am. Chem. Soc. 1981, 103, 6773-6775.

^{(3) [1-13}C]Acetate, [2-13C]acetate, [1,2-13C2]acetate, and [methyl-13C]methionine were used in a number of feeding experiments. The details of the experiments will be published elsewhere.

⁽⁴⁾ Lee, M. S.; Repeta, D. J.; Nakanishi, K.; Zagorski, M. G. J. Am. Chem. Soc. 1986, 108, 7855-7856.

⁽⁵⁾ The carbon signal assignment was made by a combination of hetero-COSY and exhaustive proton-decoupling experiments. The total proton and carbon assignment of signals in CD₂Cl₂ can be found in the supplementary material.

⁽⁶⁾ Wilson, W. B.; Collier, A. Science (Washington, D.C.) 1955, 121, 394-395

⁽⁷⁾ Chou, H. N.; Shimizu, Y.; Van Duyne, G. D.; Clardy, J. C. In *Toxic Dinoflagellates*; Anderson, D. M., White, A. W., Baden, D. G., Eds.; Elsevier/North Holland: New York, 1985, pp 305-308.

⁽⁸⁾ The ¹³C NMR sppctra were obtained in CD_2Cl_2 on a Bruker WM500 instrument at 125 MHz with SW-29 500 Hz, AQ = 0.5 s, and PW = 5 μ s. The numbers of scans (NS) are 45816 and 12404 for the samples from the methyl- and carboxyl- 13 C acetate feedings, respectively. The spectra are provided in the supplementary material along with the assignment.